



ISSN: 0975-833X

## RESEARCH ARTICLE

### CHARACTERIZATION OF WOOD ROTTING FUNGI AT MANTHA, DISTRICT JALNA (MAHARASHTRA) INDIA

\*Rajesh S. Gaikwad and Rajendra B. Kakde

Department of Botany, Swami Vivekanand Senior College, Mantha-431504 Jalna (M.S.), India

#### ARTICLE INFO

##### Article History:

Received 17<sup>th</sup> February, 2015  
Received in revised form  
23<sup>rd</sup> March, 2015  
Accepted 29<sup>th</sup> April, 2015  
Published online 31<sup>st</sup> May, 2015

##### Key words:

Macrofungi,  
Decomposers,  
Ecosystem Cleaner and  
Morphological Study

#### ABSTRACT

Occurrence of various shapes, size and colours of macrofungi reveals that the climatic conditions together with forest and garden waste provide favorable environment to these macrofungi. During the period of study from July 2014 to August 2014, many mushrooms were collected from Mantha village and at the Swami Vivekanand College campus. Study of only ten macrofungi has been emphasized on the basis of their dominance of occurrence. These are *Coprinellus micaceus*, *Guepinia spathularia*, *Irpex lacteus*, *Macrolepiota phaeodisca*, *Mycena ascendens*, *Parasola leiocephala*, *Phellinus robustus*, *Scleroderma citrinum*, *Schizopora paradoxa* and *Xylaria hypoxylon*. Morphological study of these ten wood rotting fungi was done with botanical name, common name, thallus dimension, spore dimension, spore colour, substrate and edibility.

Copyright © 2015 Rajesh S. Gaikwad and Rajendra B. Kakde. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Citation** Rajesh S. Gaikwad and Rajendra B. Kakde, 2015. "Characterization of wood rotting fungi at mantha, district jalna (Maharashtra) India", *International Journal of Current Research*, 7, (5), 16402-16406.

## INTRODUCTION

Various organisms utilize wood, of which fungi play a major role (Boddy and Watkinson 1995). As a result, wood-decaying fungi play vital role in soil formation (McFee and Stone 1966) and nutrient cycling (Dighton 1997). Some fungi invade living trees while others attack dead or down timber and grow on the forest floor. Wood rotting fungi are associated with woody host or humus rich soil. Fungi decompose wood by rotting process (Schwarze *et al.*, 2000 and Martínez *et al.*, 2005). There are three main types rots caused by fungi are white, brown and soft rot. In white rot all major constituents of wood hemicelluloses, cellulose and lignin are decomposed by fungi. On the other hand, brown rot fungi decompose hemicelluloses and cellulose and modify lignin but do not metabolise it. Yelle *et al.* (2011) reported that brown rot by *Postia placenta* results in ligninolysis, but lignin oligomers remain in wood. Soft rot fungi decompose lignin to a lesser extent than white rot fungi; wood of conifers is very resistant to soft rot (Schwarze *et al.*, 2000). In the forest, fungi decay and convert plant and animal debris into humus also carbon and nitrogen are recycled (Rossman *et al.*, 1998). Complex molecules are broken down into simple molecules, which are transported into the fungal cell.

\*Corresponding author: Rajesh S. Gaikwad,  
Department of Botany, Swami Vivekanand Senior College, Mantha-431504 Jalna (M.S.), India.

In ecosystem cycles fungi are important element, without which the food web would not be considered. Nitrogen and phosphorus are required in large quantities to biological systems, but they are not present in abundant form in the environment. Fungal action releases these elements from decaying matter and makes them available to other living organisms. Some wild edible mushrooms have been reported from South-West India (Sathe and Kulkarni, 1987). Many *Agaricus* from Andhra Pradesh have also been reported (Manoharachary and Gopal, 1991). Report of 2000 macrofungi is available from several biogeographical regions of India. But, the central India region has not been investigated extensively for mushroom flora (Kaul, 1999). Such a valuable untouched wealth of Marathwada remains neglected. As yet, this remains unexplored. Therefore, there is need to study such ecosystem cleaner. Hence, in present investigation emphasis has been given on collection of macrofungi and their detail lab study.

## MATERIALS AND METHODS

### Collection of wood rotting wood rotting fungi

From different sites of Mantha village and Swami Vivekanand Senior College Campus, ten wood rotting fungi were collected during Rainy 2014 season. Damp places, wood logs and trees were preferred for the collection of wood rotting fungi.

For collection, the paper bags were used on which the host, locality, colour of the material and date of the collection were recorded as suggested by Gilbertson and Ryvarden (1986).

### Morphological and microscopical study of macrofungi

With the help of lenses detail morphological study of macrofungi was carried out. Fruiting bodies dimensions were taken by scale.

With research microscope and ocular micrometer spore colour and spore dimension were study was carried out. The type of host and season were recorded when the sites were visited for a period of one year. Detailed microscopic examinations were made and were identified with the help of the relevant literature (Rattan, 1977; Ryvarden and Johansen, 1980; Natrajan and Kolandavelu, 1998; Lim *et al.*, 2001; Zmitrovich *et al.*, 2006; Ostry 2011).



*Coprinellus micaceus* (Bull.) Vilgalys, Hopple & Jacq. Johnson



*Guepinia spathularia* (Schweinitz)



*Irpex lacteus* (Fr.) Fr.



*Macrolepiota phaeodisca* Bellù



*Mycena ascendus* (Lasch) M. Geest.



*Parasola leiocephala* (P. D. Orton) Redhead, Vilgalys & Hopple



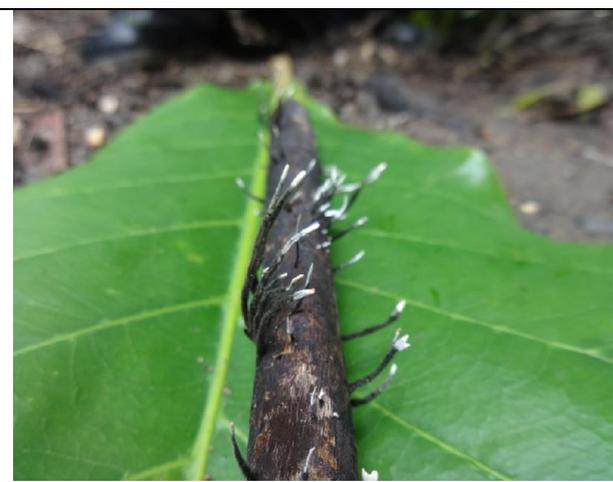
*Phellinus robustus* (L.) Quel.



*Schizopora paradoxa* (Schrad. : Fr.) Donk



*Scleroderma citrinum* Pers.



*Xylaria hypoxylon* (L.) Grev.

**Table 1. Morphological and microscopical study of wood rotting fungi**

Botanical name	Common name	Thallus Dimension	Substrate/ Host	Season	Edible	Spore dimension	Spore color
<i>Coprinellus micaceus</i> (Bull.) Vilgalys, Hopple & Jacq. Johnson -	Glistening inkcap	Cap: Covered at first in tiny white granules, the egg-shaped caps become bell shaped and lose their salt-like grains of velum as they age. The caps are typically 3 to 5cm tall and of similar diameter when they open out. The cap colour is ochre-brown, hygrophanous. Gills: The gills are white, turning purple-brown and then blackening. Stem: The white stem is brownish at the base, typically 3 to 6mm in diameter and 5 to 9cm tall.	Debris of Agriculture wastes	Rainy 2014	Yes (those grown at only natural sites)	Ellipsoidal to shield-shaped, smooth, 8-10 x 3.5-6µm; with a central germ pore.	Dark brown
<i>Guepinia spathularia</i> (Schweinitz)	Fan shaped jelly fungus	Fruit body: Gelatinous and rubbery; yellow; greasy, with curved, blunt or pointed horns rarely forked at the tips; 2 to 12mm tall and typically 1 to 2mm dia.	Babool door slit	Rainy 2014	Yes	Ellipsoidal to sausage-shaped, smooth, 7-10 x 2.5-4µm;	white or very pale yellow coloured.

Continue.....

<i>Irpex lacteus</i> (Fr.) Fr.	Toothed Polypore	Fruiting body: A spreading patch of creamish pore surface with 2-3 pores per mm, tooth-like rather than poroid, except at margin; develop shelflike edges or kidney-shaped caps.	Wood log of <i>Annona squamosa</i>	Rainy 2014	No	Spores 5-7 x 2-3 $\mu$ ; smooth; elliptical to subcylindric	White
<i>Macrolepiota phaeodisca</i> Bellù	--	Cap: Convex and pale brown area near the crown that breaks into scales. The cap cuticle is floccose, covered with irregular concentric rings of flakes and most pronounced near the margin, diameter at maturity ranges between 4 and 8cm. Gills: The broad, crowded gills of this mushroom are white or pale cream and free, terminating close to the stipe.	Humus rich soil	Rainy 2014	Yes	Ellipsoidal, smooth, thick-walled; typically 10 x 5 $\mu$ ; with a small germ pore.	White
<i>Mycena ascendens</i> (Lasch) M. Geest.	Frosty bonnet	Cap: 2-4.5 mm across, hemispherical, conical, translucent-striate, shallowly sulcate, white, glabrescent. Stipe 5-30 mm long, filiform, straight to flexuous, equal, greyish-hyaline, puberulous, glabrescent with age	Wood log of <i>Azadirachta indica</i>	Rainy 2014	Unknown	7.0-9.5 x 3.0-6.0 $\mu$ , broadly smooth, thin-walled, smooth,	Amyloid
<i>Parasola leiocephala</i> (P. D. Orton)	Pleated Inkcap	Cap: 1 to 2cm diam., heavily ribbed, convex and flat. A distinctive central 'eye' contrasts with the rest of the pale grey cap. Gills: The white gills turn grey and then black; they are free of the stem Stem: Up to 6cm long, but only 4 or 5mm in diameter, and very fragile; white.	Grown at crust of brick industry	Rainy 2014	No	Irregularly heart shaped and ellipsoidal, 8.0-11.5 x 7.0-10 x 5-7.5 $\mu$ , with a small germ pore.	Black
<i>Phellinus robustus</i> (L.) Quel.	--	Fruit body: Woody, hoof-shaped, brown to black and crusty upper surface, rusty brown interior, firm and woody, attached by a broad lateral base, 60 cm wide, 5cm thick, margin entire, reddish brown. Pore surface plane, often creviced, with a sterile border 2-5 mm wide, reddish brown to dark brown; pores stratose, small, 5-7 per mm	On babool tree	Rainy 2014	Yes	Basidiospores globose to subglobose, 8 x 7 $\mu$ , smooth.	Hyaline
<i>Schizophora paradoxa</i> (Schrad. : Fr.) Donk	Split Porecrust	Fruit body: Patches are developed, ochraceous yellow with creamy-white margins, fertile surface covered in blunt teeth up to 4mm long.	Wood log of <i>Annona squamosa</i>	Rainy 2014	No	Cylindrical sausage-shaped, smooth, 8-10 x 3-3.5 $\mu$ .	White
<i>Scleroderma citrinum</i> Pers.	Earthball	Fruit body: 7cm across and 6cm tall, rounded, Fruitbody stemless and attached to the ground by white mycelial threads. White. Inside the earthball the spore mass is present.	Humus rich soil	Rainy 2014	No	Spherical, spiny, 8-13 $\mu$ diameter when fully mature.	Brownish
<i>Xylaria hypoxylon</i> (L.) Grev.	Candlesnuff Fungus	Fruit body: Small, upright 2 to 8mm in diameter at the base and typically 3 to 5cm tall. Spikes and antlers like. Initially black and finely downy near the sterile base and white with conidia towards the tips.	Dry woody stick of <i>Annona squamosa</i>	Rainy 2014	No	Bean shaped, smooth, 11-15 x 4-6 $\mu$ . Black.	Black

## RESULTS

All ten specimens collected were observed with their external and internal characters of basidiocarp and spores. Macroscopical and microscopical study of wood rotting fungi has been given in Table 1. Lakhanpal (1996) studied the occurrence of wood rotting fungi on substrate like, wood, litter and soil indicates that their vital role in ecosystem. Mushrooms from Waster Ghats were recorded by Pradeep *et al.* (1998). From Punjab plains, Atri *et al.* (2000) has carried out detail taxonomic study of *Agaricus* species. There are 41,000 species of Mushrooms in the world, of which 850 species occur in India (Deshmukh, 2004). Similarly, Sharma and Samota (2006) reported the growth of wood rotting fungi on humus, dung, saw dust, gardens, roadsides and deserts. Kakde and Gaikwad (2014) reported some polypores, puff fungi, bracket fungi and fleshy fungi, grown on humus rich soil, leaf litter and woody debris of *Annona squamosa*, *Annona reticulata* and *Azadirachta indica*. In present study puff fungus and bracket fungus are recorded from same substrates. Hadawoo (2010) reported some edible mushrooms from genera like *Agaricus*, *Coprinus*, *Cyathus*, *Lycoperdon*, *Schizophyllum*, *Daldinia*, *Polyporus* and *Ganoderma* from Amravati region of Maharashtra.

Through this study we are just reporting diversity of wood rotting fungi from our region for the first time. Many wood rotting fungi from Mantha region of Jalna district of Maharashtra, have high ability to remediate contaminated soil and leaf litter, wood logs with lignin degradation. Further study is required to explore the use of these organisms for the waste management and production of neutraceuticals and therapeutics.

## Acknowledgement

Authors are thankful to Principal, Swami Vivekanand Senior College Mantha, Jalna (M.S.) for providing research facilities.

## REFERENCES

Atri, N.S., Kaur, A. and Saini, S.S. 2000. Taxonomic studies on *Agaricus* from Punjab plains. *Indian J. Mushroom*, 18, 6-14.

Boddy, L. and Watkinson, S.C. 1995. Wood decomposition, higher fungi, and their role in nutrient redistribution. *Canadian Journal of Botany*, 73 (Suppl. 1): S1377-S1383.

Deshmukh, S.K. 2004. Biodiversity of tropical basidiomycetes as sources of novel secondary metabolites. In *Microbiology and Biotechnology for Sustainable Development* (ed. Jain PC), CBS Publishers and Distributors, New Delhi, pp 121-140.

Dighton, J. 1997. Nutrient cycling by saprotrophic fungi in terrestrial habitats. In: Wicklow DT, Söderstrom BE (eds) *The Mycota*, vol. 4, Environmental and microbial relationships. Springer, Berlin, p. 271-279.

Gilbertson, R.L. and Ryvardeen, L. 1986. North American polypores., Vol. 1, pp.433. Fungiflora, Oslo, Norway.

Kakde, R.B. and Gaikwad, R.S. 2014. Diversity of wood decay fungi at Mantha, Jalna (M.S.) India. *Biosci. Disc.*, 5(2) 230-236.

Kaul, T.N. 1999. Introduction to mushroom science. Oxford and IBH Publi. Co. Pvt. Ltd. N. Delhi, 198p.

Lakhanpal, T.N. 1996. *Mushrooms of Indian Boletaceae Vol-I. Studies in Cryptogamic Botany* (ed. Mukharji KG), APH Publishing Coporation Delhi.

Lim, Y.W., Kim, H.Y. and Jung, H.S. 2001. The Aphyllophorales of Mungyong Saejae. *Mycobiology*, 28(3) 142-148.

Manoharachary, C. and Gopal, K.V. 1991. Mycofloristics of Agaricales from Andhra Pradesh. In: *Indian Mushrooms 1991*, Kerala Agriculture University, Vellanikkara, India.

Martínez, Á.T., Speranza, M., Ruiz-Dueñas, F.J., Ferreira, P., Camarero, S., Guillén, F., Martínez, M.J., Gutiérrez, A. and del Río, J.C. 2005. Biodegradation of lignocellulosics: microbial, chemical, and enzymatic aspects of the fungal attack of lignin. *International Microbiology*, 8, 195-204.

McFee, W.W., Stone, E.L., 1966. The persistence of decaying wood in the humus layers of northern forests. *Soil Science Society of America Journal*, 30, 513-516

Natarajan, K. and Kolandavelu, K. 1998. Resupinate Aphyllophorales of Tamil Nadu, India. Centre for advanced study in Botany, University of Madras, 133pp.

Ostry, M.E., Anderson, N.A. and O'Brien, J.G. 2011. Field guide to common macrofungi in eastern forests and their ecosystem functions. Gen. Tech. Rep. NRS-79. Newtown Square, PA: U.S. Department of Agriculture, Forest Service, Northern Research Station. 82 p.

Pradeep, C.K., Virinda, K.B., Mathew, S. and Abraham, T.K. 1998. The genus *Volvariella* in Kerala state, India. *Mushroom Res.*, 53-62.

Rattan, S.S. 1977. The Resupinate Aphyllophorales of the North Western Himalayas. J. Cramer, In der A.R. K.G. FL-9490 Vaduz, Germany, 427pp

Rossmann, A., Tulloss, R.E., O'Dell, T.E. and Thorn, R.G. 1998. All Taxa Biodiversity Inventory of Fungi in a Costa Rican Conservation Area. Parkway Publishers, Inc., Boone, NC. 195 p.

Ryvardeen, L. and Johansen, J. 1980. A preliminary Polypore Flora of East Africa, Fungiflora. Oslo, Norway, 636pp.

Sathe, A.V. and Kulkarni, S.M. 1987. A checklist of wild edible mushrooms from South-West India. In: *Indian Mushroom Science II*. Kaul TN and Kapur BM eds, Regional Res. Lab., Jammu, 411-413.

Schwarze, F.W.M.R., Engels, J. and Mattheck, C. 2000. Fungal strategies of wood decay in trees. Springer Verlag, Berlin Heidelberg New York.

Sarma, S.S. and Samota, R.K. 2006. Tapping into India's Mushroom biodiversity; Identification, Conservation and Domestication of wild Mushroom flora. Compedium of lectures-*Emerging Areas in Mushroom diversity, production and post harvest development*. Organized by Mushroom Research Laboratory, Deptt. of Plant Pathology. Indira Gandhi Agriculture University, Raipur. 69-84.

Yelle, D.J., Wei, D., Ralph, J. and Hammel, K.E. 2011. Multidimensional NMR analysis reveals truncated lignin structures in wood decayed by the brown rot basidiomycete *Postia placenta*. *Environmental Microbiology*, 13, 1091-1100.

Zmitrovich, I.V., Malysheva, V.F. and Wjacheslav, A.S. 2006. A new morphological arrangement of the Polyporales. I. Phanerochaetinae. *Mycena*, 6, 4-56.