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RESEARCH ARTICLE

SEROPREVALENCE OF ANAPLASMA sp. IN SHEEP (OVIS ARIES) BY ELISA IN PESHAWAR, PAKISTAN

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ABSTRACT

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Key words: Anaplasmosis, Epidemiology, MSP-5, Peshawar. In the present study, sero-prevalence of anaplasma sp. in sheep, *Ovisaries* (L) was done from January-May, 2012 in Peshawar, Pakistan. The information concerning anaplasmosis in sheep is scare. For this purpose, 376 serum samples were obtained randomly from4 different breeds of sheep, fromdifferent areas of Peshawar, and an indirect ELISA using recombinant MSP-5 as antigen of *Anaplasmamarginale* (T), was performed. Totally, 92/376 (24.47%) of the overall sheep sera were positive for antibodies against *A. marginale*. In 4 breeds of sheep, (i.e. Balkhai, Watanai, Punjabai and Turkai) Turkai were found highly infected i.e. 27/376 (7.20%). This is the first record of *A. marginal* infection in Sheep in Peshawar, Pakistan, which is very high. This research should be useful in epidemiological applications.

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INTRODUCTION

Sheep, Ovisaries (L) is one of the initial animals, domesticated for agricultural purposes; it is raised for meat, (hogget or mutton, lamb) milk and fleece production. These quadru-pedal ruminant mammals are members of the order Artiodactyla, the even-toed ungulates typically kept as livestock. It hasgreat economic potential because of their early maturity and high fertility as well as their adaptability to moist environment (Ademosun, 1988). However, the benefits derived are too low from the expected due chiefly to low productivity. Numerous factors are involved in this low productivity, in which the major one is disease (Akerejola et al., 1979).Diseases caused by heamoparasites are most apparent. These heamoparasites are parasites found in the blood of mammals in which A. marginale is also include. Ticks are biological vectors of Anaplasmasp.; tick, mammalian or bird hosts with persistent Anaplasmasp. infection can serve as reservoir of infection naturally. Anaplasmasp. is intracellular, gram-negative bacteria and representatives of the order Rickettsiales classified into Rickettsiaceae and Anaplasmataceae families (Dumler et al., 2001). The tick vector distribution is the factor influencing the transmission of tick-borne diseases (Bazarusanga et al., 2007a). However, for A. marginale, mechanical transmission

*Corresponding author:Muhammad Kashif Department of Zoology, Hazara University, Mansehra, Pakistan. through contaminated hypodermic needles and biting flies plays an important role (Potgieterand Stoltsz, 2004). Erythrocytes are phagocyted by reticulo-endothelial cells during infection. Animals may die older than 2 years due to the infection (Kocan et al., 2003). Nevertheless, concerning ovine anaplasmosis, little information is available, in despite of the expressive number of sheep, goat and expansion of small ruminant herds in this country. Diagnosis of anaplasmosis in small ruminants mainly based in the identification of the rickettsia in stained blood smears. However, below 0.1% rickettsemias in chronic carriers are not detected by this method (Palmer, 1992). Serological assays, based on Major Surface Protein 5 (MSP-5) of A. marginale have been successfully used, for the detection of antibodies against Anaplasma sp. (Strik et al., 2007). In this study, we observed for the first time sero-prevalence of Anaplasma sp., in different breeds of sheep using an indirect ELISA based on MSP5 recombinant of A. marginale, in Peshawar, Pakistan. This research should be particularly useful for epidemiological applications such as prevalence studies, awareness, education, research and control programs in this region.

MATERIALS AND METHODS

Samples collection

Conveniently, 376 blood sampling was collected from the overall sheep population of different areas of Peshawar from

Table 1. Sex wise positive samples of sheepanaplasmosis by indirect Enzyme Linked Immunosorbent Assay (iELISA) from January to May, 2012 from Peshawar, Pakistan

S. No.	Sheep Breeds	Age					
5 . NO.		n^1	Adult n ²	Positive*(%)	Young n ³	Positive**(%)	
1	Balkhai	124	78	14 (17.94)	46	9 (19.56)	
2	Watanai	97	70	18 (25.71)	27	8 (29.62)	
3	Punjabai	76	42	8 (19.04)	34	8 (23.52)	
4	Turkai	81	36	10 (27.77)	45	17 (37.77)	

 \mathbf{n}^1 : total numbers of samples examined; \mathbf{n}^2 : total numbers of male samples examined

 \mathbf{n}^3 : total numbers of female samples examined; (*) (**) represents positive samples for *A.marginale* of male and female respectively

 Table 2. Age wise positive samples of sheepanaplasmosis by indirect Enzyme Linked Immunosorbent Assay (iELISA) from January to May, 2012 from Peshawar, Pakistan

S. No.	Sheep Breeds	Sex					
		n ¹	Male n ²	Positive*(%)	Female n ³	Positive**(%)	
1	Balkhai	124	93	17 (18.28)	31	6 (19.35)	
2	Watanai	97	71	21 (29.57)	26	5 (19.23)	
3	Punjabai	76	58	12 (20.68)	18	4 (22.22)	
4	Turkai	81	65	22 (33.85)	14	7 (50.00)	

 \mathbf{n}^{1} : total numbers of samples examined; \mathbf{n}^{2} : total numbers of adult samples examined

 n^3 : total numbers of young samples examined; (*) (**) represent positive samples for *A.marginale* of adult and young respectively

January to May 2012. About 5 ml blood samples were collected from the jugular vein of each sheep with a sterile hypodermic syringe into an evacuated tube containing gel breed, age, and sexwere noted. The blood sample was then centrifuge for 5 minutes at 12000 rpm to separate serum and stored at -35 C until further use. The SVANOVIR[®]A. *marginale*-Ab ELISA kit (Svanova Biotech AB, Uppsala, Sweden) was used for the diagnosis of specific antibodies against *A. marginale* in bovine serum samples. The kit procedure was based on the Indirect Enzyme Linked Immunosorbent Assay (Indirect ELISA). The whole procedure was done according to the protocol given with the kit.

Protocol for Indirect Enzyme Linked Immunosorbent Assay (iELISA)

All reagents were equilibrated to room temperature 18 to 25 °C before use. Pre-dilution of control and samples 1/40 in PBStween buffer (e.g., 10 µl sample in 390 µl of PBS-tween buffer). Hundred micro liter of pre-diluted serum sample was added to selected wells. The plate was then sealed and incubated at 37 °C for 30 minutes. The plate was rinse 4 times with PBS-tween buffer. Hundred micro liter of conjugate dilution was added to each well and then sealed the plate and incubate on 37 °C for 30 minutes. Again, and clot activator. Some information like the plate was rinse 4 times with PBStween buffer. Hundred micro liter substrate solution was added to each well and then incubated for 30 minute at room temperature (18 to 25 °C). Hundred micro liter of stop solution was added to each well and mixed thoroughly. The Optical Density (OD) of the controls and sample was measured at 405 nm in a micro-plate photometer (BIOTEK Instruments Inc., Winooski, Vermont, U.S.A.). Mean OD values were calculated for each of the control and samples.

Data analysis

The following formula was used for the Percent Positivity (PP):PP= [(sample OD $\times 100$)/Mean positive control OD]

Interpretation of the results

The calculatedPercent Positivity(PP) if less than 25%, the sample was consider as negative and if PP was equal or more than 25%, then the sample was consider as positive.

RESULTS

There were overall 92 (24.47%) positive and 284 (75.53%) negative blood samples for A.marginaleof sheep breeds were: Balkhai—18.55% (23/124),Watanai—26.80% (26/97),Punjabai-21.05% (16/76) and Turkai 35.80% (29/81) (Figure 2). Sex wise, in total 124 Balkhai samples, 17/93(18.28%) male positive and 6/31 (19.35%) were female positive. In total 97 Watanai samples, 21/71 (29.58%) male positive and 5/26 (19.23%) were female positive. In Punjabai 76 samples, 12/58 (20.69%) male positive and 4/18 (22.22%) were female positive. In Turkai 81, 22/65 (33.85%) male positive and 7/14 (50.00%) were female positive (Table 1). Age vise in total 124 Balkhai, they were 9/46 (19.56%) young positive and 14/78 (17.95%) were adult positive. In total 97 Watanai, there were 8/27 (29.63%) young positive and 18/70 (25.71%) were adult

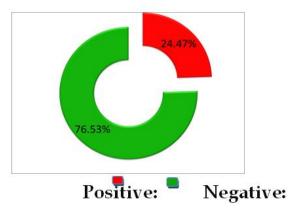
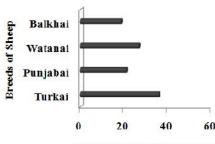


Fig. 1. Number of positive (24.47%) and negative (76.53%) samples of sheep (*Ovisaries*) of *A.marginale*by indirect Enzyme Linked Immunosorbent Assay (iELISA) from January to May, 2012 from Peshawar, Pakistan



Percentage of Positve Samples

Fig. 2. Breeds wise percentage of positive samples of sheep(O. aries) of A.marginaleby indirect Enzyme Linked Immunosorbent Assay (iELISA) from January to May, 2012 from Peshawar, Pakistan

positive. In total 76 Punjabai, there were 8/34 (23.53%) young positive and 8/42 (19.05%) were adult positive. In total 81 Turkai, they were 17/45 (37.77%) young positive and 10/36 (27.77%) were adult positive (Table2).

DISCUSSION

The research on sheep anaplasmosis (A. marginale) is rare and little literature is available. The frequencies in the four positivity of sheep anaplasmosisin this research were (24.47%) which is very low as compared to the prevalence of seropositive sheep found by Hornok et al. (2007) (99.4%) in Hungry and high as compared to the prevalence of seropositive sheep found by Cabral et al. (2009) (8.92%). Seroprevalence were found byRamos et al. (2008) (16.17%) in Ibimirim county, semi-arid region of Pernambuco State, Brazilusing monoclonal antibody ANAF16C1 and De La Fuente et al. (2005) (75.0%), in sicily, Italy, using competitive ELISA, based on recombinant MSP-5 of A. marginale. The low sero-prevalence rate in this research work can be the cause of low tick vector populationin Peshawararea. However, some ticks were also observed in sheep during blood samples collection. This result represents the first description of antibodies for Anaplasmasp. in sheep from Peshawar, Pakistan. Further studies are require to know the epidemiology of Anaplasmasp. infection in sheep, in Pakistan, particularly to define which species is involved, possible impacts and vectors in animal production and in public health.

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