



RESEARCH ARTICLE

ANTIBACTERIAL ACTIVITY OF EUCALYPTUS OIL AGAINST CLINICAL ISOLATES OF STAPHYLOCOCCUS AUREUS

*Noorul Aneesa and Dr. Gopinath, P.

Department of Microbiology, Saveetha Dental College, Chennai, India

ARTICLE INFO

Article History:

Received 28th March, 2017

Received in revised form

14th April, 2017

Accepted 12th May, 2017

Published online 20th June, 2017

Key words:

Staphylococcus aureus,
MIC. Eucalyptus oil.

Copyright©2017, Noorul Aneesa and Dr. Gopinath, P. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Noorul Aneesa and Dr. Gopinath, P. 2017. "Antibacterial activity of eucalyptus oil against clinical isolates of staphylococcus aureus", *International Journal of Current Research*, 9, (06), 51850-51852.

ABSTRACT

Staphylococcus aureus is one of the important bacterial pathogen causing a wide spectrum of infections. Eucalyptus species are well known as medicinal plants due to their biological and pharmacological properties. The aim of the present study was to determine the antibacterial activity of eucalyptus oil against clinical isolates of *Staphylococcus aureus*. The MIC of eucalyptus oil was appeared to be 0.06% for *S. aureus*. The eucalyptus oil is found to have antibacterial activity against *S. aureus*. However, its irritant properties has been evaluated before it is formulated for medicinal purpose.

INTRODUCTION

Staphylococcus aureus is one of the important bacterial pathogen causing a wide spectrum of infections (Arciola *et al.*, 2001). Many studies have been conducted to explain the structures and pathogenic mechanisms by which *S. aureus* is able to cause serious infections (O'Neill *et al.*, 2007). The ability of *S. aureus* to produce biofilm enables this organism to withstand the host immune response and is considered to be the cause of many chronic or persistent infections, as the biofilm creation protects bacteria from phagocytosis and antimicrobial agents (Foster, 2005). Another concern related to this pathogen is increasing resistance to oxacillin and many other antibiotics, but also circulation of multidrug resistant isolates within the hospital environment (Martín-López *et al.*, 2002). Staphylococcal pathogenesis is multifactorial, involving a combination of adherence and biofilm formation (Klug *et al.*, 2003). Eucalyptus is one of the very important and most widely planted genera across the world (Akin *et al.*, 2010). It is a tall, evergreen tree, native to Australia and Tasmania, successfully introduced worldwide, now extensively planted in many other countries (Mubita *et al.*, 2008). It was introduced in Algeria in 1854 by Ramel (Boulekbache-Makhlouf *et al.*, 2010). Eucalyptus species are well known as medicinal plants due to their biological and pharmacological properties. In the international pharmacopeia, the most important and represented species, however, is *Eucalyptus globulus*

(*E. globulus*) which is the main furnisher of essential oils. These essential oils have different applications as anesthetic, anodyne, antiseptic, astringent, deodorant, diaphoretic, disinfectant, expectorant, febrifuge, fumigant, hemostat, inhalant, insect repellent, preventitive, rubefacient, sedative yet stimulant, vermifuge, for a folk remedy for abscess, arthritis, asthma, boils, bronchitis, burns, cancer, diabetes, diarrhea, diphtheria, dysentery, encephalitis, enteritis, erysipelas, fever, flu, inflammation, laryngalgia, laryngitis, leprosy, malaria, mastitis, miasma, pharyngitis, phthisis, rhinitis, sores, sore throat, spasms, trachalgia, worms, and wounds. Sometimes their demand is also high in the soap and cosmetic industries (Bajaj, 1995). Thus, the aim of the present study was to determine the antibacterial activity of eucalyptus oil against clinical isolates of *Staphylococcus aureus*.

MATERIALS AND METHODS

Bacterial isolates

A total of 20 clinical isolates of *S. aureus* were collected from different clinical specimens of patients attending Saveetha Medical Collage and hospital. They were processed for a battery of standard biochemical tests and confirmed. Isolates were preserved in semisolid trypticase soy medium and stored at 4°C until further use.

Antibiotic susceptibility test: Antibiotic susceptibility testing was determined for these isolates to the following antibiotics

*Corresponding author: Noorul Aneesa

Department of Microbiology, Saveetha Dental College, Chennai, India

such as penicillin, erythromycin, clindamycin, ciprofloxacin, tetracycline, cotrimoxazole and linezolid. These antibiotics were procured from Himedia, Mumbai. This was performed by Kirby-bauer disc diffusion method as per CLSI guidelines (Clinical Laboratory Standards Institution, 2015).

Detection of antibacterial activity of eucalyptus oil against clinical isolates of *S. aureus*

Anti-bacterial activity of eucalyptus oil was tested against *S. aureus* isolates by minimum inhibitory concentration method. Mueller Hinton broth was supplemented with 0.002% (V/V) tween 80 (HiMedia, Mumbai) to enhance the dispersion of the essential oil. Agar dilution method was performed to attain the different concentrations of essential oils such as 0.03%, 0.06%, 0.125%, 0.25%, 0.5%, 1% and 2% in Mueller Hinton Agar (MHA).

Media containing various concentrations of essential oils were poured over the sterile petridishes and allowed to dry. Media without essential oil was served as control plate. Spot inoculation of 0.5 McFarland standard turbidity adjusted isolates were made on the plates and incubated at 37°C for overnight. The lowest concentration of the essential oils that completely inhibited the growth of isolates was considered as MIC (Gopinath Prakasam *et al.*, 2014).

RESULTS

Sample wise distribution of clinical isolates of *S. aureus*

Of 20 clinical isolates of *S. aureus*, 8/20 (40%) were obtained from pus, 6/20 (30%) were from wound, 4/20 (20%) and 2/20 (10%) were from blood and sputum respectively (Figure 1).

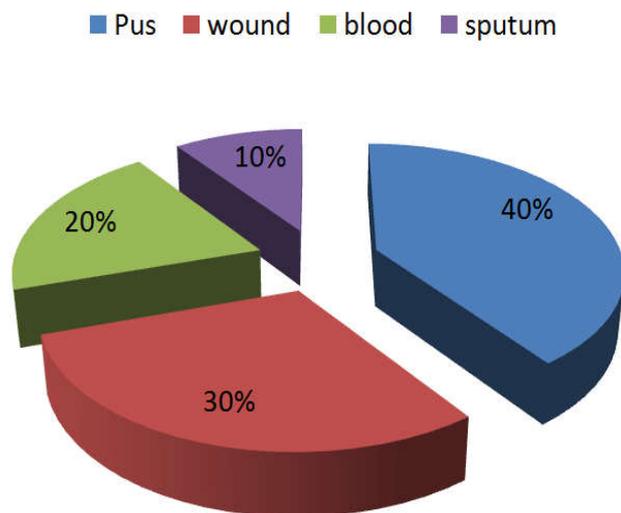


Figure 1. Pie chart showing the sample wise distribution of *S. aureus*

Antibiotic susceptibility pattern

We have observed a varied pattern of sensitivity among one *S. aureus* isolates. There was complete resistance observed for penicillin(100%), 9/20(45%)isolates were shown to the resistant to erythromycin,6/20(30%) were to cotrimoxazole,4/20(20%)were to linezolid followed by 3/20(15%) were resistant to ciprofloxacin and clindamycin respectively (Table 1) (Figure 2).

Table 1. Results of antibiotic susceptibility pattern of *S. aureus*

Antibiotics	Sensitive (%)	Intermediate (%)	Resistant (%)
Penicillin	0	0	20(100)
Erythromycin	14(70)	4(20)	2(10)
Clindamycin	15(75)	2(10)	3(15)
Ciprofloxacin	9(45)	8(40)	3(15)
Tetracyclin	14(70)	4(20)	2(10)
Cotrimoxazole	10(50)	4(20)	6(30)
Linezolid	10(50)	6(30)	4(20)

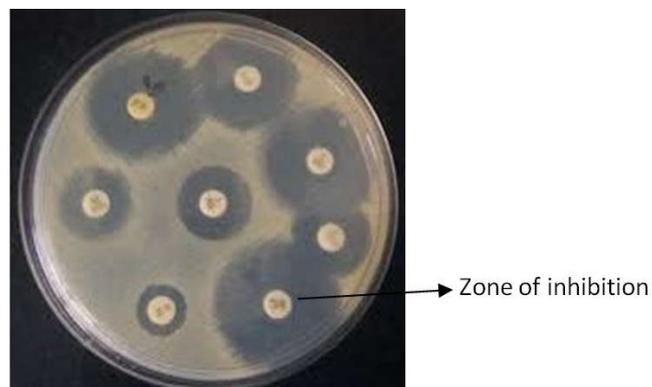


Figure 2. Representative picture showing antibiotic sensitivity pattern of *S. aureus*

Result of antibacterial activity of eucalyptus oil against clinical isolates of *S. aureus*

We have observed that, clinical isolates of *S. aureus* were inhibited from 0.06-1% of eucalyptus oil. The MIC of eucalyptus oil was appeared to be 0.06% for *S. aureus*.

Dilutions of eucalyptus oil %	0.03 %	0.06 %	0.125 %	0.25 %	0.5%	1%	2%
No. of organisms	0	7 (35%)	7 (35%)	1 (5%)	2 (10%)	3 (15%)	0

DISCUSSION

Study conducted by Prakasam *et al.* from Chennai in 2014 demonstrated that, *Acinetobacter* strains were inhibited from 0.06 to 0.25%, 0.25-1% and 0.125-1% for clove, peppermint and eucalyptus oils respectively. In clove oil, 14/50 (28%) isolates were inhibited at 0.06%, 25/50 (50%) at 0.125% and 11/50 (22%) at 0.25% of clove oil. In peppermint oil, 34/50 (68%) isolates were inhibited at 0.25%, 12/50 (24%) and 4/50 (8%) were at 0.5% and 1% concentrations of peppermint oil respectively. In eucalyptus oils, 10/50 (20%) isolates were inhibited at 0.125%, 18/50 (36%) at 0.25%, 16/50 (32%) and 6/50 (12%) were at 0.5% and 1% respectively. Thus, the MIC of clove oil was found to be 0.06%, 0.25% for peppermint oil and 0.125% for eucalyptus oil (Gopinath Prakasam *et al.*, 2014). In contrast, in our study, we used eucalyptus oil against *S. aureus* isolates. 35% of isolates were inhibited at 0.06%, 35% were at 0.125%, 5% were at 0.25%, 10% were at 0.5% and 1% were at 15% of essential oil. Thus, the MIC of eucalyptus oil against *S. aureus* was found to be 0.25%.

Conclusion

The eucalyptus oil is found to have antibacterial activity against *S. aureus*. However, its irritant properties has been evaluated before it is formulated for medicinal purpose. Due to the extended drug resistance in *S. aureus*, it can be used as an alternative medicine.

REFERENCES

- Akin, M., Aktumsek, A., Nostro, A. 2010. Antibacterial activity and composition of the essential oils of *Eucalyptus camaldulensis* Dehn and *Myrtus communis* L. growing in Northern Cyprus. *Afr J Biotechnol.*, 9:531–535.
- Arciola, C.R., Baldassarri, L., Montanaro, L. 2001. Presence of icaA and icaD genes and slime production in a collection of staphylococcal strains from catheter-associated infections. *The Journal of Clinical Microbiology*, 39 (6). 2151–2156.
- Bajaj, Y.P.S. 1995. Medicinal and aromatic plants. Berlin, Heidelberg, New York: Springer Edition, Volume 8, Biotechnology in agriculture and forestry; pp. 194–196.
- Boulekbache-Makhlouf, L., Meudec, E., Chibane, M., Mazauric, J.P., Slimani, S., Henry, M., et al. 2010. Analysis by high-performance liquid chromatography diode array detection mass spectrometry of phenolic compounds in fruit of *Eucalyptus globulus* cultivated in Algeria. *J Agric Food Chem.*, 58(24):12615–12624. [PubMed]
- Clinical Laboratory Standards Institution: Performance standards for antimicrobial susceptibility testing. In NCCLS approved standard M2-A8. Wayne, PA USA: CLSI; 2015
- Foster, T.J. 2005. Immune evasion by staphylococci. *Nat Rev Microbiol.*, 3(12). 948–958.
- Gopinath Prakasam, Manju Bhashini, Lakshmi Priya, Srivani Ramesh, S. 2014. In-vitro antibacterial activity of some essential oils against clinical isolates of *Acinetobacter baumannii*. *Indian J Med Microbiol.*, 32:90-91.
- Klug, D., Wallet, F., Kacet, S., Courcol, R.J. 2003. Involvement of adherence and adhesion Staphylococcus epidermidis genes in pacemaker lead-associated infections. *The Journal of Clinical Microbiology*, 41 (7); 3348–3350.
- Martín-López, J.V., Pérez-Roth, E., Claverie-Martín, F., Díez Gil, O., Batista, N., Morales, M., Méndez-Alvarez, S. 2002. Detection of Staphylococcus aureus Clinical Isolates Harboring the ica Gene Cluster Needed for Biofilm Establishment. *The Journal of Clinical Microbiology*, 40(4). 1569–1570.
- Mubita, C., Syakalima, M., Chisenga, C., Munyeme, M., Bwalya, M., Chifumpa, G., et al. 2008. Antibigrams of faecal *Escherichia coli* and *Enterococci* species isolated from pastoralist cattle in the interface areas of the Kafue basin in Zambia. *Veterinarski Arhiv*, 78(2):179–185.
- O'Neill, E., Pozzi, C., Houston, P., Smyth, D., Humphreys, H., Robinson, D.A., O'Gara, J.P. 2007. Association between methicillin susceptibility and biofilm regulation in Staphylococcus aureus isolates from device-related infections. *The Journal of Clinical Microbiology*, 45(5). 1379–1388.
