



RESEARCH ARTICLE

A STUDY ON HISTOLOGICAL ALTERNATIONS IN GILL AND LIVER OF *Heteropneustes fossilis* UPON EXPOSURE TO HERBICIDE 2, 4 DICHOLOPHENOXYACETIC ACID

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ABSTRACT

This study was carried out to evaluate the effects of 2, 4 D on gill and liver tissue of *Heteropneustes fossilis* in order to understand its toxicity. The experimental fishes were exposed to two sublethal concentrations 3.00 ppm (1/10<sup>th</sup> of LC<sub>50</sub>) and 0.60 ppm (1/50<sup>th</sup> of LC<sub>50</sub>) doses of 2, 4 D for duration of 30 days. Histopathological analysis showed damage to liver and changes in gill architecture which were characterized by vacuolation, hepatic necrosis, disruption of gill lamellae. Alternation or change in gill structure could impair oxygen intake which eventually might cause death of the fish. The damage in liver tissue might be an indication of poor physiological activity as different important enzymes are secreted by it. Hence the application of herbicides in agricultural field and aquatic body should be monitored as it can reach humans through food chain.

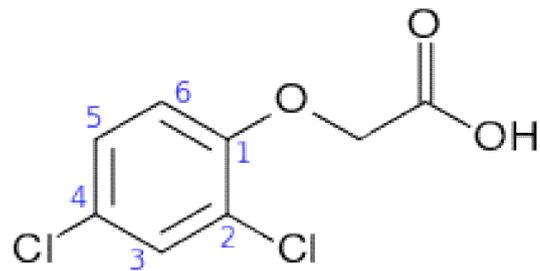
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INTRODUCTION

The effect of different chemicals has serious impact on the health of humans as well as animals of both aquatic and terrestrial habitat. Herbicides are chemicals which are commonly used to control undesirable herbs and vegetation. It is applied in water bodies to control aquatic weeds. It can also enter the aquatic ecosystem through surface runoff. However, high concentration of herbicides could lead to significant losses of aquatic vegetation and algal biomass, which could ultimately hamper the aquatic fauna. As pesticides are not fully selective, it is found from Environment Toxicity studies that these substances affect non-target species in the environment (Rao, 2006). When the concentration of herbicide exceeds tolerant level it causes damage to important vital organs and reduces the survival, growth and the reproduction performance of aquatic organisms (Rahaman et al., 2002). Fish is good indicator of aquatic contamination because its biochemical stress response is quite similar to those found in mammals (Mishra and Shukla, 2003). 2, 4 Dichlorophenoxyacetic acid, usually referred to as 2, 4 D, is a chlorinated phenoxy herbicide, often used in agricultural fields. Commercially it can be found in various forms such as acids, amines, various salts and ester derivatives, which vary in chemical properties and toxicity.

It is used for weed control of rice, wheat, maize, aquatic weeds and also employed for post-emergence foliar spray (Farah et al., 2004).



Chemical structure of 2, 4-D

The intensive and widespread use of 2, 4 D exhibit deleterious physiological, behavioural, neurological, immunological, histological and hematological alternations as reported by many observers at different times (Tuschl and Schwab, 2003; Stürtz et al., 2008; Uyanikgil et al., 2009; Kubrak et al., 2013) and has led to the emergence of herbicide resistant weeds (Watanabe et al., 1997; Bradberry et al., 2000; Toyama et al., 2003). Thus, this study was undertaken to establish the effect of sublethal concentrations of 2, 4 Dichlorophenoxyacetic acid (a herbicide) on histopathological changes in gills and liver of catfish, *Heteropneustes fossilis*.

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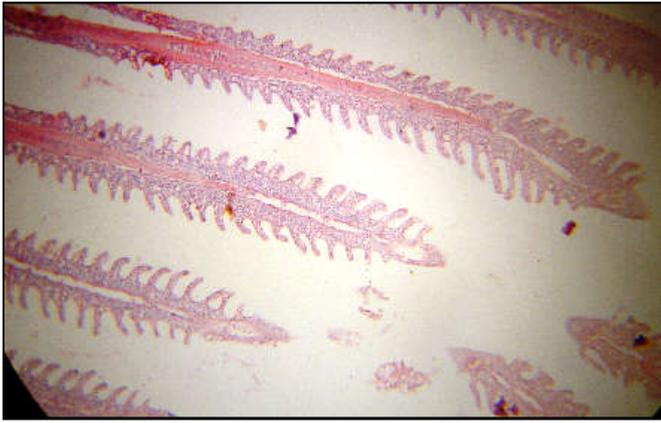


Fig. 1. Gill structure of normal control fish. (Magnification X100)

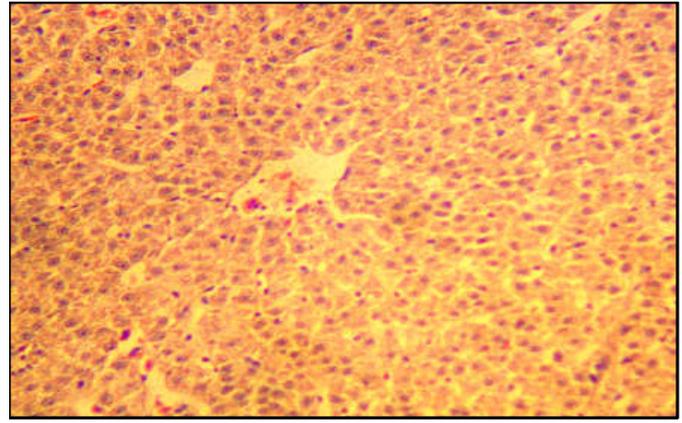


Fig. 4. Liver of normal control fish. (Magnification X100)

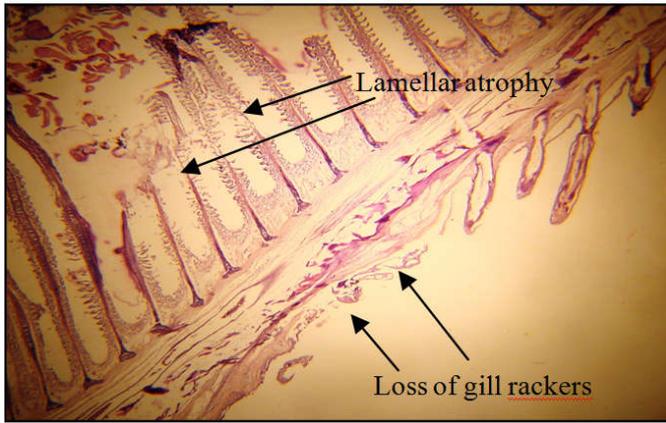


Fig. 2. 3.00 ppm (1/10<sup>th</sup> of LC<sub>50</sub>) 2, 4 D treated Gill. (Magnification X100)

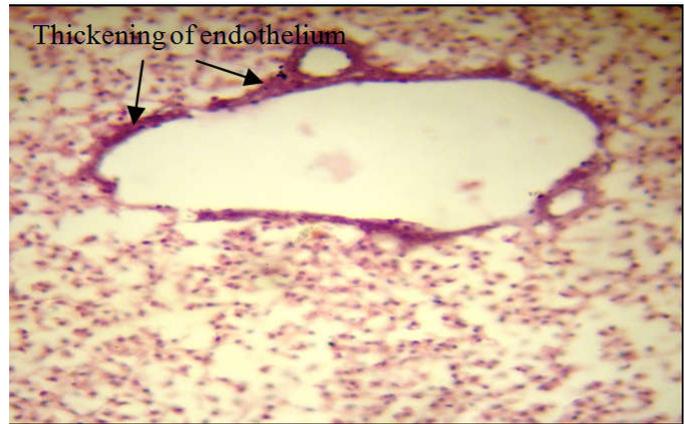


Fig. 5. 3.00 ppm (1/10<sup>th</sup> of LC<sub>50</sub>) 2, 4 D treated Liver. (Magnification X400)

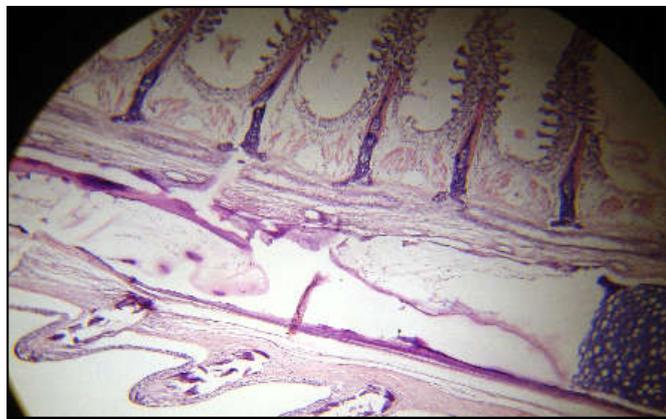


Fig. 3. 0.60 ppm (1/50<sup>th</sup> of LC<sub>50</sub>) 2, 4 D treated Gill. (Magnification X400)

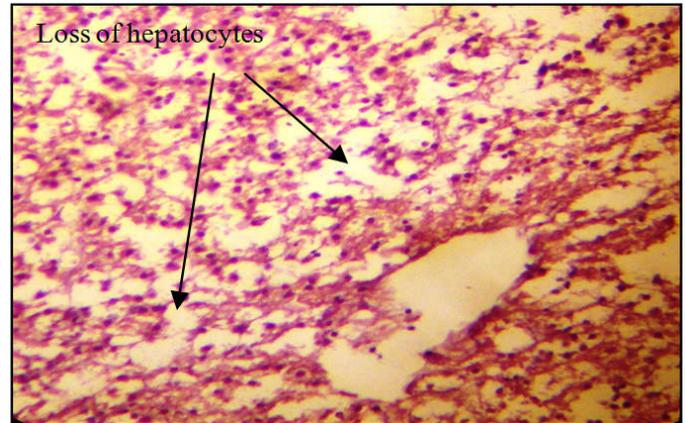


Fig 6: 0.60 ppm (1/50<sup>th</sup> of LC<sub>50</sub>) 2, 4 D treated Liver. (Magnification X100)

**MATERIALS AND METHODS**

The experimental fish, *Heteropneustes fossilis* weighing about 10 gm and length of about 11 cm were purchased from local fish market of Dhubri town, Assam. Prior to the start of the experiment, the fishes were acclimatized for about a week in an aquarium under suitable laboratory conditions. Sublethal dose of 2, 4 D (3.00 ppm i.e., 1/10<sup>th</sup> of LC<sub>50</sub> and 0.60 ppm i.e., 1/50<sup>th</sup> of LC<sub>50</sub>) were administered separately in the experimental fishes for a period of one month (i.e., 30 days). The experiments were carried out in three different aquaria, one of which was left untreated as normal control group and the other two each containing the different treated group.

The fishes were grouped in three different groups each containing 5 fishes:

- Group-I represent normal control group,
- Group-II represent 2, 4 D (3.00 ppm) treated group and
- Group-III represent 2, 4 D (0.60 ppm) treated group.

**Histological method**

Fishes were collected from each group randomly after the exposure of 30days and dissected to collect the liver and gills. The specimens were taken in normal saline solution and fixed in 10% formalin. It was then hydrated in graded alcohol, then cleared in xylene and finally embedded in molten paraffin

block. Using rotary microtome, the specimens were sectioned at 4µm thickness. Standard H & E method was used for staining and finally mounted in DPX. The slides as prepared were examined under microscope for histopathological analysis.

## RESULTS

### Histopathological changes

**Gills:** The gill is made up of filaments of primary lamellae from which secondary lamellae arise. Gill arches and gill rays are clearly observed in Group-I fishes (Fig. 1). Comparing with the normal control set, it was found that the fish treated with different sublethal dose of 2, 4 D shows various histopathological changes. The gill of 3.00 ppm 2, 4 D treated group shows degenerative changes with loss of gill rakers as well as atrophy of lamellar structure in the gill filaments (Fig. 2). Mild congestions of blood vessels were observed in the primary lamellae of 0.60 ppm 2, 4 D treated *H.fossilis*. The epithelial layer was also seen to be swollen (Fig.3).

**Liver:** No histological changes were observed in the liver tissue of normal control group (Group-I) of experimental fish. The liver appears as continuous mass of hepatic cells or hepatocytes having cord like arrangement with the central vein (Fig. 4). Result of the present study demonstrated that the sublethal concentration of 2, 4 D produces large histopathological alterations in the liver of treated fishes. Focal and diffuse area of haemorrhage, extensive necrosis with loss of hepatocytes was observed. Thickening of the endothelium of the central vein was also observed in experimental fishes treated with 3.00 ppm concentration of 2, 4 D (Fig.6). Whereas in lower dose of 2, 4 D (0.60 ppm) treated fishes, the cytoplasm of liver tissue showed increased granularity with disintegration of hepatocytes. Focal areas of necrosis and pyknosis of the nuclei were observed in the liver tissue (Fig.5). Moreover in some areas complete loss of hepatic structures were seen.

## DISCUSSION

Histopathological anomalies due to contamination of different toxic chemicals have been revealed by many researchers. Moreover the frequency of pathological changes increases with increase in the concentration of toxicants. Fish gill is an organ with a large surface area, highly sensitive to the effect of toxicant (Reiser *et al.*, 2010). The work of Anusuya and Hemalatha, (2015) showed that gills of *Channa striatus* treated with 2, 4 D (100mg/l) has hypertrophy, fusion and shortening of lamellae and necrosis in gill structure. Samanta *et al.*, (2016) also observed similar findings in glyphosate (17.20 mg/l) exposed *H.fossilis*. Literature review showed that histopathological changes such as fusion of secondary lamellae accompanied by hypertrophy or hyperplasia of primary filaments occurred in *Tilapia zilli* treated with aluminium (Hadi and Alwan, 2012). However in 2, 4 D amine treated *C.gariepinus* for 96hr showed mild desquamation of the epithelium, vacuolization and lifting of epithelial layer as observed by Makinde *et al.*, (2015). The liver of bluegill showed rapid shrinkage of parenchymal cells and loss of vacuolation as examined by Cope *et al.*, (1970). 15 mg/kg of 2, 4 D treated rat showed disrupted hepatic cords at many areas and upon exposure to 150mg/l 2, 4 D, the liver showed pycnotic nucleus, congestion of hepatocytes and necrotic cells

at the end of 4 weeks of treatment (Tayeb *et al.*, 2010). Histopathological alternations were also observed in *Rhamdia quelen* liver upon exposure to 700mg/l of 2, 4 D for 96 h (Cataneo *et al.*, 2008). The parenchymal cell of liver underwent rapid shrinkage and loss of vacuolation as observed by Cope *et al.*, (1970) upon exposure to 2, 4 D. 96 hr exposure to acute concentration of 2, 4 amine showed hepatitis (Makinde *et al.*, 2015). Degeneration and disintegration in most cytoplasmic contents, necrosis along with pyknosis and rupture of hepatocytes were observed in the work of Pandey and Dubey, (2015) on *H.fossilis* upon exposure to pentachlorophenol (PCP) for 21 days. Deka and Mahanta (2012) were also observed pyknotic nuclei in liver of malathion treated *H.fossilis*. Thus the present investigation showed somewhat similar resemblance with the above findings. The study showed that 2, 4 D at different sublethal concentration causes significant histomorphological changes in the liver and gill of *H.fossilis*. The histopathological changes were seen to be more pronounced in sublethal concentration at higher dose of 2, 4 D than lower dose level. Moreover, the induced histopathological changes in the gills may lead to several physiological stresses in the fish. Hence, we conclude from the results of the present study that the low concentration of 2, 4 D used in this research shows potent histopathological effects while the 2, 4 D toxicity was found to be significant at high dose level causing degenerative changes in tissue architecture.

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## REFERENCES

- Anusuya J. and Hemalatha S. 2015. A study on histopathological impact of 2, 4 Dichlorophenoxyacetic acid on gills and liver of *Channa striatus*. *Int. J. Modn. Res. Revs.*, Vol. 3 (5), 669-673.
- Bradberry, S.M., Watt, B.E., Proudfoot, A.T. and Vale, J.A. 2000. Mechanisms of toxicity, clinical features and management of acute chlorophenoxy herbicide poisoning: a review. *J. Toxicol. Clin. Toxicol.*, 38, 111-122.
- Cattaneo R., Loro V.L., Spanevello R., Silveira F.A., Luzb L. B., Miron D.S., Fonseca M.B., Moraes B.S. and Clasen B. 2008. "Metabolic and histological parameters of silver catfish (*Rhamdia quelen*) exposed to commercial formulation of 2,4-dichlorophenoxyacetic acid (2,4-D) herbicide." *Pesticide Biochemistry and Physiology*, 92, 133-137.
- Cope O.B., Wood E. M. and Wallen G.H. 1970. Some chronic effects of 2, 4-D on the Bluegill (*Lepomis macrochirus*). *Trans. Amer. Fish. Soc.*, Vol.99 (1), 1-12.
- Deka S. and Mahanta R. 2012. A study on the effect of organophosphorus pesticide Malathion on Hepato-Renal and Reproductive organs of *Heteropneustes fossilis* (Bloch), *The Science Probe*, vol 1 (1), 1-13.
- Farah M.A., Ateeq B., Ali M.N., Sabir R. & Ahmad W. 2004. Studies on lethal concentrations and toxicity stress of some xenobiotics on aquatic orga-nisms. *Chemosph.*, 55:257-265.
- Hadi A.A and Alwan S.F. 2012. Histopathological changes in gills, liver and kidney of fresh water fish, *Tilapia zillii*,

- exposed to aluminum. *Int. J. of Pharm and Life Sci.*, Vol.3 (11), pp.2071-2081.
- Kubrak, O.I., Atamaniuk, T.M., Husak, V.V. and Lushchak, V.I. 2013. Transient effects of 2,4 dichlorophenoxyacetic acid (2,4-D) exposure on some metabolic and free radical processes in goldfish white muscle. *Food Chem. Toxicol.*, 59:356-361.
- Makinde G.E.O., Olaifa F. E. and Banjo O.T. 2015. Acute Toxicity and Histopathological Changes in Gill and Liver of Catfish (*Clarias Gariepinus*) Juvenile Exposed to 2, 4-D Amine (Herbex D Sl®). *J. of Bio., Agri and Healthcare.*, Vol.5 (4), 145-149.
- Mishra R. and Shukla S. P. 2003. Endosulfan effects on muscle malate dehydrogenase of the freshwater catfish *Clarias batrachus*. *Ecotox. Environ. Safe.*, 56, 425-433.
- Pandey A.K and Dubey S. 2015. Histological changes in liver and kidney of cat fish, *Heteropneustes fossilis*, exposed to pentachlorophenol PCP. *Plant Archives*, Vol. 15 (2), 1117-1120.
- Rahman M.Z., Hossain Z., Mellah M.F.A. and Ahmed G. U. 2002. Effects of diazinon 60EC on *Anabus testudineus*, *Channa punctatus* and *Barbades gomonotus Naga*. *The ICLARM Quarterly*, 25, 8-11.
- Rao J.V. 2006. Toxic effects of novel organophosphorus insecticide (RPR-V) on certain biochemical parameters of euryhaline fish *Oreochromis mossambicus*. *Pest. Biochem. Physiol.*, 86, 78-84.
- Reiser S., Schroeder J.P., Wuertz S., Kloas W. and Hanel R. 2010. Histological and physiological alterations in juvenile turbot (*Psetta maxima*, L.) exposed to sublethal concentrations of ozone produced oxidant. *Aquaculture*. 307, 157 – 164.
- Samanta P., Mukherjee A.K., Pal S., Kole D. and Ghosh A.R. 2016. Toxic effects of glyphosate-based herbicide, Excel Mera 71 on gill, liver, and kidney of *Heteropneustes fossilis* under laboratory and field conditions. *J. of Microscopy and Ultrastruc.*, 4, 147–155
- Stürtz N., Ricardo P.D., Jahn G.A., Duffard R., Maria A. and de Duffarda E. 2008. Effect of 2,4-dichlorophenoxyacetic acid on rat maternal behavior. *Toxicology*, 247, 73-79.
- Tayeb W., Nakbi A., Trabelsi M., Attia N., Miled A. and Hammami M. 2010. Hepatotoxicity induced by sub-acute exposure of rats to 2,4-Dichlorophenoxyacetic acid based herbicide “Désormone lourde”. *Journal of Hazardous Materials*, 180, 225–233.
- Toyama K., Bae C.H., Kang J.G., Lim Y.P., Adachi T., Riu K.Z., Song P.S. and Lee H.Y. 2003. Production of herbicide-tolerant *zoysiagrass* by Agrobacterium-mediated transformation. *Mol. Cells*, 31, 19–27.
- Tuschl H. and Schwab C. 2003. Cytotoxic effects of the herbicide 2,4- dichlorophenoxyacetic acid in HepG2 cells. *Food Chem. Toxicol.*, 41, 385-393.
- Uyanıkgil Y., Yalcıncay M., Ateş U., Bakaa M. and Karakişi H. 2009. Effects of 2,4- dichlorophenoxyacetic acid formulation on medulla spinalis of *Poecilia reticulata*: A histopathological study. *Chemosphere*, 76, 1386-1391.
- Watanabe H.M., Ismail Z. and NaiKin Ho. 1997. Response to 2, 4-D resistant biotype of *Fimbristylis miliacea* (L.) Vahl. to 2, 4-D methylamine and its distribution in the Muda Plain, Peninsular Malaysia. *J. Weed Sci. Technol.*, 42, 240–249.

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