



RESEARCH ARTICLE

ANTIOXIDANT ACTIVITY OF NORTH INDIAN FRUIT ANANASCOMOSUS (PINEAPPLE) AND VITISVINIFERA (GRAPES) USING 1,1-DIPHENYL 1-2-PICRYLHYDRAZYL METHOD (DPPH)

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INTRODUCTION

The effect of consuming food and beverages rich in polyphenols in terms of preventing diseases such as cancer and coronary diseases (Aqil *et al.*, 2006; Beyer, 1994; Braca *et al.*, 2002) is quite significant. Polyphenols can also reduce damage to DNA and production of free radicals in the body (Cakmak and Marschner, 1992; Cao *et al.*, 1996; Samman, 1996; Duh *et al.*, 1999). Many of the flavonoids found in grape juice, such as catechin, epicatechin, quercetin, and anthocyanins are known to have antioxidant, anti-inflammatory, and platelet inhibitory effects, as well as for being able to reduce LDL oxidation and oxidative damage to DNA, both in vitro and in animal studies (Halliwell, 1994; Jensen, 1978; Kanatt *et al.*, 2007; Koleva *et al.*, 2002) Colour is the most important attribute used, along with other variables, as an indicator of the quality of grape juice. This characteristic is directly dependent on the phenolic composition of the juice and the anthocyanin's present in the grape skin (Kalt and Kushad, 2001; Ansari *et al.*, 2005; NaikSeema *et al.*, 2008). The anthocyanins participate in many reactions that promote changes in the color of grape

products, mainly through copigmentation and formation of polymeric pigments (Mendis *et al.*, 2005; SajithaRajan *et al.*, 2007; Ramakrishna *et al.*, 2008).

MATERIALS AND METHODS

All solvents and chemicals used were analytical /BDR grade. DPPH was obtained from March, Mumbai, India. Fruits were purchased from local market in Nagpur Maharashtra India.

Plant material

Healthy and fresh grapes and pineapple were selected and purchased from the total market of Nagpur.

Preparation of Samples

Fruits were cleaned under running tap water excessive water was drained off. The fruits were cut into small pieces and subjected to size reduction using kitchen blender with a kitchen mixer to get a thick paste, and kept at 20°C for further analysis.

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Preparation of extract

250mg of plant samples was extracted with 10ml of 100% methanol and left it overnight, Next day filtered with Whitman filter paper and make up the volume up to 25ml with 100 ml ethanol.

RESULTS AND DISCUSSION

Anti-oxidant activity

Free radical scavenging activity by DPPH method

Free radical scavenging potential of fruits extract at different concentrations was tested by DPPH method and the results are presented in Table 1. It can be seen that the two fruits exhibition varying degrees of in vitro total antioxidant capacity (Pi-Jen Tsai et al., 2002; Singh et al., 2002). The average antioxidant of grapes and pine apple were 50.22% and 54.99 % RSA with methanolic DPPH solution respectively

Table 1. Free radical scavenging activity by DPPH method

Deferent	Optical Density		RSA %	
Concentration of DPPH	grapes	pineapple	grapes	pineapple
(ppm)				
25	0.083	0.080	51.74	53.48
50	0.081	0.074	52.90	56.97
100	0.088	0.076	48.83	55.81
250	0.103	0.080	40.11	53.48
500	0.073	0.077	57.55	55.23
Blank	OD=			
0.1723	Total DPPH Value		50.22%	54.99%

Conclusion

The value of the polyphenolic content is high. The results are a direct measure of soluble polyphenolic content the test is so sensitive that no interfering color is present at the dilutions used. A simple and rapid method for evaluating antioxidant activity was developed. The antioxidant activity against DPPH method is of the useful in characterizing the properties antioxidant activity of the substance found in fruits.

REFERENCES

- Ansari N M, Houlihan L, Hussain B and pieroni A. 2005. antioxidant activity of five fruits traditionally consumed by south Asian migrants in Bradford Yorkshire (UK) *Phytother. Res.*, 19,907-911
- Aqil F, Ahmed I, Mehmood Z 2006. Antioxidant and free radical scavenging properties of twelve traditionally used Indian medicinal plants. *Turk. J. Biol.*, 30:177-183.
- Beyer RE. 1994. The role of ascorbate in antioxidant protection of biomembranes: interaction with vit-E and coenzyme. *Q. J. Bioen. Biomemb.*, 24:349-358.
- Braca A, Sortino C, Politi M 2002. Antioxidant activity of flavonoids from *Licanialicaniaeflora*. *J. Ethnopharmacol.*, 79: 379-381.
- Cakmak I, Marschner H 1992. Magnesium deficiency and high light intensity enhance activities of superoxide dismutase, ascorbate peroxidase and glutathione reductase in bean leaves. *Plant Physiol.*, 98:1222-1227.
- Cao G, Sofic ER, Prior RL 1996. Antioxidant capacity of tea and common vegetables. *J. Agric. Food Chem.*, 44: 3426-3431. Cook NC,
- Duh PD, Tu YY, Yen GC 1999. Antioxidants activity of aqueous extract of Harnjyur (*Chrysanthemum morifolium* Ramat). *Lebensmwiss Technol.*, 32: 269-277.
- Halliwell B 1994. Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? *Lancet*, 344: 721-724.
- Jensen A 1978. Chlorophyll and carotenoids. In: Hallebust JA, Craigie JS.(eds). *Handbook of Physiochemical and Biochemical Methods*. Cambridge University Press, Cambridge, UK, pp. 5-70.
- Kalt W and Kushad MM 2001. The role of oxidative stress and antioxidants in plant and human health introduction to the colloquium. *Horticulture science* PriorR L and Cao G 2000 antioxidant phytochemicals in fruits and fruits . *Diet and health implications, Horticulture Science*, 35: 588-592
- Kanatt SR, Chander R, Sharma A 2007. Antioxidant potential of mint (*Mentha spicata* L.) in radiation- processed lamb meat. *Food Chem.*, 100: 451-458.
- Koleva II, Van Beek TA, Linseen JPH, de Groot A, Evstatiieva LN 2002. Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. *Phytochem. Anal.*, 13:8
- Mendis E Rajapaksa N, Kim SK 2005. Antioxidant properties of radical scavenging peptide purified from enzymatically prepared fish skain gelatin hydroysate j. of agric. *Food Chem.*, 53; 581-587
- NaikSeema M, Jayaprakasha G.K, Singh RP 2008. Antioxidant activity of custard apple (*annona squamosal*) peel and seed extract, *J. of Food Science Technol.*, 45(4) 349-352
- Pi-Jen Tsai, John McIntosh, Philip Pearce, Blake Camden, Brian R Jordan 2002. Anthocyanin and antioxidant capacity in Roselle (*Hibiscus Sabdariffa* L.) extract, 35(351-356)
- Ramakrishna, B. V. and Jayaprakasha, G. K. and Jena, B. S. and Singh, R. P. 2008. Antioxidant activities of froselle (*Hibiscus sabdariffa*) calyces and fruit extracts. *Journal of Food Science and Technology*, 45 (3). pp. 223-227
- SajithaRajan S, Shilpastheeh L, Kishormohan T C, Murugan K 2007. Value of ethnic foods in meeting antioxidants needs: the wild plant connection, *J. of Food Science Technology*, 44(4)394-396
- Samman, S. 1996. Flavonoids-chemistry, metabolism, cardioprotective effects and dietary sources. *Nutr. Biochem.*, 7:66- 76.
- Singh RP, Chidambara Murthy KN, Jayaprakasha GK. 2002. Studies on the antioxidant activity of pomegranate (*Punicagranatum*) peel and seed extracts using in vitro models. *J.Agric Food Chem.*, Jan 2;50(1):81-6.
