

Available online at http://www.journalcra.com

International Journal of Current Research Vol. 8, Issue, 02, pp.25983-25986, February, 2016 INTERNATIONAL JOURNAL OF CURRENT RESEARCH

REVIEW ARTICLE

ROLE OF LACTIC ACID BACTERIA AS PROBIOTIC: A REVIEW

Geeta Devi and *Geetika Mehta

Department of Genetics and Plant Breeding, CCS Haryana Agricultural University, Hisar- 125004, Haryana

ARTICLE INFO

ABSTRACT

Article History: Received 16th November, 2015 Received in revised form 22nd December, 2015 Accepted 27th January, 2016 Published online 14th February, 2016

Key words:

Lactic Acid Bacteria, Probiotic, Bacteriocins, Health. Lactic acid bacteria are industrially important organisms recognized for their fermentative ability as well as their health and nutritional benefits. There is an increasing interest in these species to reveal the many possible health benefits associated with them. The actions of LAB are species and strain specific, and depend on the amount of bacteria available in the gastrointestinal tract. However, products with or processed with LAB are accepted as a natural way to preserve food and promote health. This paper aimed to review the recent data in regard to the role of probiotic LAB in food industries, in the immunomodulation in the gastrointestinal tract, and there application in its health benefits.

Copyright © 2016 Geeta Devi and Geetika Mehta. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Geeta Devi and Geetika Mehta, 2016. "Role of lactic acid bacteria as Probiotic: A review", *International Journal of Current Research*, 8, (02), 25983-25986.

INTRODUCTION

Lactic acid bacteria (LAB) are characterized as Gram-positive cocci or rods, non-aerobic but aerotolerant, able to ferment carbohydrates for energy and lactic acid production. pathway The metabolic from glucose may be homofermentative or heterofermentative. In the first case two molecules of lactate are generated (as in Streptococcus and Lactococcus), and in the second, lactate, ethanol and carbon dioxide as in Leuconosto. Lactic acid bacteria are also able to produce small organic substances that contribute with aroma and give specific organoleptic attributes to the products (Caplice and Fitzgerald et al., 1999). The microorganisms are found in milk, meat and fermented products, as well as in fermented vegetables and beverages inhibiting the growth of pathogenic and deteriorating microorganisms, maintaining the nutritive quality and improving the shelf life of foods. They have also been used as flavor and texture producers. Lactic acid bacteria include various major genera: Lactobacillus, Lactococcus, Corynebacterium, Enterococcus, Lactosphaera, Leuconostoc, Melissococcus, Oenococcus, Pediococcus, Streptococcus, Tetragenococcus, Vagococcus and Weissella. Other genera are: Aerococcus, Microbacterium,

*Corresponding author: Geetika Mehta,

Department of Genetics and Plant Breeding, CCS Haryana Agricultural University, Hisar-125004, Haryana.

Propionibacterium and Bifidobacterium (Carr et al., 2002) are some of the most common species (Garrity, 1984., Dellaglio et al., 1994), and some strains are recognized as probiotics (Fuller, 1989., Parada et al., 2003). Sugar fermentation followed by a reduction in pH due to the production of lactic and other organic acids is an important factor for the inhibition of growth of undesired microorganisms. The low pH makes organic acids liposoluble, them to break through the cell membrane and reach the cytoplasm of pathogens (Haller *et al.*, 2001). The competition for essential nutrients, accumulation of D-amino-acids and diminution of the oxiredutive potential also contribute to their inhibitory effect. LAB are usually known as safe (GRAS), and have an important role in the preservation of foods and fermented products. They can be used as natural competitive microbiota or as specific starter cultures under controlled conditions (Cintas et al., 2001). Some of these bacteria produce antagonistic substances, called bacteriocins, which in small amounts are very active against pathogens (Klaenhammer et al., 1994; Moreno et al., 2006) Lactic acid bacteria (LAB) occur naturally in several raw materials like milk, meat and flour used to produce foods (Rodriguez et al., 2000). LAB are used as natural or selected starters in food fermentations in which they perform acidification due to production of lactic and acetic acids flavour. Protection of food from spoilage and pathogenic microorganisms by LAB is through producing organic acids, hydrogen peroxide, diacethyl (Messens and De Vugst., 2002), antifungial compounds such

as fatty acids (Corsetti *et al.*, 1998) or phenullactic acid (Lavermicocca *et al.*, 2000) and/or bacteriocins (De Vugst and Vandamme., 1994). LAB play an important role in food fermentation as the products obtains with their aid are characterized by hygienic safety, storage stability and attractive sensory property. Many bacteria of different taxonomic branches and in various habitats produce antimicrobial substances that are active against other bacteria. Both Gram negative and Gram positive bacteria produce bacteriocins. Bacteriocins are proteinaceous antibacterial compounds, which constitute a heterologous subgroup of ribosomally synthesized antimicrobial peptides (De Vugst and Vandamme., 1994).

Lactic acid bacteria as probiotics

Lactic acid bacteria were referred to as probiotics in scientific literature by Lilley and Stillwell (1965). However probiotic took on a different terminology when Sperti (1971) used the term probiotic to describe tissue extracts that stimulated microbial growth. Parker (1974) redefined it as organisms and substances that contribute to the intestinal microbial balance. The most recent and accurate description of probiotics was undertaken by Fuller (1989) who redefined it as a live microbial feed supplement beneficial to the host (man or animal) by improving the microbial balance within its body. Another recent definition was by Schrezenmeir and De Vrese (2001) who defined probiotics as viable microbial food supplements which beneficially influence the health of the host. The gastrointestinal tract contains food in different stages of digestion, digestive ferments, liquids and solid waste. Within the gut are also wide ranges of microbes that may be either harmful or beneficial. The beneficial ones assist in the breakdown of food while they also manufacters vitamins essential to the body, breaking down and destroying some toxic chemicals that may have been ingested with the food. Under both healthy and sick conditions, several differnt types of bacteria compete or fight with each other to establish dominance in the warm and moist environment of the alimentary canal that serves as an ecosystem for their survival and propagation.

The average human large intestine harbors over 400 different special of bacteria with a total population far outnumbering even the number of human cells in the body. Under ideal conditions of health and diet, the different strains of bacteria on microflora compete and check the excessive number of any one strain. Healthy condition can be achieved if a balance is maintened between the good and bad bacteria in the ratio of 85 percent to 15 percent. Oral supplement of diet with viable Lactbacillus acidophilus of human origin, which is bile resistant, led to a significant decline of three different fecal enzymes (Goldin and Gorbach et al., 1977). This bacterial decrease in the fecal bacterial enzyme activity observed in both humans and rats included beta glucuronidase, azoreductase and nitroreductase. All these enzymes catalyse the conversion of procarcinogens to proximal carcinogens in the large bowel leading to colon cancer. Lactic acid bacteria including Lactobacillus, leuconostoc, lactococcus, pediococcus and Bifidobacterium are found throughout the gastrointestinal tract. The predominant population of lactic acid bacteria in the upper

gastrointestinal tract is the Lactobacillus species which may colonize the mucosal surface of the duodenum as well as the stomach. Lactobacillus and Bifidobacterium spp. are prominent members of the commensal intestinal flora and are the commoly studied probiotics bacteria. They cause reduced lactose intolerance alleviation of some diarrhoeas, lowered blood cholesterol, increased immune response and prevention of cancer (Marteau and Ramband, 1993, 1996 Gilliland, 1996; Salminen et al., 1998). The selection criteria for probiotic LAB include: human origin, safety, viability activity in delivery vehicles, resistance to acid and bile, adherence to gut epithelial tissue ability to colonise the gastro intestinal tract, production of antimicrobial substances, ability to stimulate a host immune response and the ability to influence metabolic activities such as vitamin production, cholesterol assimulation and lactose activity (Salminen et al., 1996). Conway (1996) and Fuller (1989) listed the following organisms as species used in probiotic preparation: L. acidophilus, L. casei, L. casei subsp. rhamnosus, Lactobacillus fermentum, Lactobacillus reuteri Lactococcus lactis subsp lactis, L. cremoris, L. bulgaricus, L. Streptococcus thermophilus, Enterococcus plantarum, faecium, Enterococcus faecalis, B. bifidum, B. infantis, B. adolescentis, B. longum, B. breve.

Probiotics benefit in the gastro intestinal tract and Immune system

Certain LAB species are found not only as components of the human intestinal microflora but also of the man made ecosystem present in fermented food. That is why milks containing viable LAB are known to be beneficial to health acting as prophylaxis against intestinal infections. Thus many investigators have evaluated the effect of yoghurt on the immuno response of animals and humans. Many studies have been conducted on their effect on the incidence and duration of various types of diarrhoe (Isolauri, 2001; Bhatnagar et al., 1998). LAB can be effective in preventing gastrointestinal disorders and in the recovery from diarrhoea of miscellaneous causes (Marteau et al., 2001). A decrease in the severty and duration of persistent diarrhoea has been reported with LAB (Bhatnagar et al., 1998). Guandalini et al., (2000) also reported that the administration of Lactobacillus rhamnosus GG to 287 children aged 1- 36 months with acute diarrhoea significantly reduced the duration in infected children by rotavirus compared with those receiving placebo. Administration of Lb rhamnosus GG also shortened the duration of the hospital stay.

Application of LAB

Lactic acid bacteria are industrially important organisms recognized for their fermentative ability as well as their health and nutritional benefits. Species used for food fermentations belong to the genera *Lactococcus, Streptococcus, Pediococcus, Leuconostoc, Lactobacillus,* and the newly recognized *Corynbacterium.* These organisms have been isolated from grains, green plants, dairy and meat products, fermenting vegetables, and the mucosal surfaces of animals.Once used to retard spoilage and preserve foods through natural fermentations, they have found commercial applications as starter cultures in the dairy, baking, meat, vegetable, and alcoholic beverages industries. They produce various compounds such as organic acids, diacetyl, hydrogen peroxide, and bacteriocins or bactericidal proteins during lactic fermentations not only are these components desirable for their effects on food taste, smell, color and texture, but they also inhibit undesirable microflora. Hence, lactic acid bacteria and their products give fermented foods distinctive flavors, textures, and aromas while preventing spoilage, extending shelf-life, and inhibiting pathogenic organisms. The preservative action of starter culture in food and beverage systems is attributed to the Combined action of a range of antimicrobial metabolites produced during the fermentation process. These include many organic acids such as lactic, acetic and propionic acids produced as end products which provide an acidic environment unfavourable for the growth of many pathogenic and spoilage microorganisms.

REFERENCES

- Atrih, A., Rekhif, N., Moir, A. J. G., Lebrihi, A. and Lefebvre, G. 2001. Mode of action, purification and amino acid sequence of plantaricin C19, an anti-*Listeria* bacteriocin produced by *Lactobacillus plantarum* C19. *International Journal of Food Microbiology*, 68: 93-109.
- Audisio, M. C., Terzolo, H. R. and Apella, M. C. 2005. Bacteriocin from Honeybee Beebread *Enterococcus avium*, Active against *Listeria monocytogenes*. *Applied and Environmantal Microbiology*, 71: 3373-3375.
- Beasley, S. S. and Saris, P. E. J. 2004. Nisin-Producing Lactococcus lactis Strain isolated from Human Milk. Applied and Environmental Microbiology, 70: 5051-5053.
- Bendali, F., Gaillard-Martinie, B., Hebraud, M. and Sadoun, D. 2008. Kinetic of production and mode of action of the *Lactobacillus paracasei* subsp. *paracasei* anti-listerial bacteriocin, an Algerian isolate. *LWT-Food Science and Technology*, 41:1784-1792.
- Bhatnagar, S, Singh, K D, Sazawal, S, Saxena, SK, BHAN, MK 1998. Efficacity of milk versus yoghurt offered as part of a mixed diet in acute noncholera diarrhoea among malnourished children. *Journal of paediatrics*, 132: 999-1003.
- Bhunia, A. K., Johnson, M. C. and Ray, B. 1987. Direct detection of an antimicrobial peptide of *Pediococcus* acidilactici in sodium dodecyl sulphate-polyacrylamide gel electrophoresis. *Journal of Industrial Microbiology*, 2: 319-322.
- Bredholt, S., Nesbakken, T. and Holck, A. 2001. Industrial application of an antilisterial strain of *Lactobacillus sakei* as a protective culture and its effect on the sensory acceptability of cooked, sliced, vacuum-packaged meats. *International Journal of Food Microbiology*, 66:191-196.
- Bruna, C.G., Carolina, T.E., Izabel, C.V.P., Ana, L.C.D., Felis, G.E., Leonardo, A.S., Bernadette, D.G.M.F. and De Martinis, E.C.P. 2008. Prevalence and characterization of *Enterococcus* spp. isolated from Brazilian foods. *Food Microbiology*, 25: 668–675.
- Casaus, P., Nillsen, T., Cintas, L. M., Nes, I. F., Hernandez, P. E. and Holo, H. 1997. Enterocin B, a new bacteriocin from *Enterococcus faecium* T136 which can act synergistically with enterocin A. *Microbiology*, 143: 2287-2294.
- Cintas, L.M, Rodriguez, J.M, Fernandez, M.F, Sletten, K., Nes I.F, Hernandez, P.E., Holo, H. 1995. Isolation and

characterization of pediocin L50, a new bacteriocin from *pediococcus acidilactici* with broad inhibitory spectrum. *Applied Environmental Microbiology61:2643-2648*.

- De Kwaadsteniet, M., Todorov, S. D., Konoetze, H. and Dicks, L. M. T. 2005. Characterization of a 3944 Da bacteriocin produced by *Enterococcus mundtii* ST15, with activity against Gram positive and Gram negative bacteria. *International Journal of Food Microbiology*, 105: 433-444.
- De Vuyst, L., Callewaert, R. and Pot, B. 1996. Characterization of the antagonistic activity of *Lactobacillus amylovovorus* DCE 471 and large scale isolation activity of its bacamylovorin L471. *Systematic and Applied Microbiology*, 19: 9-20.
- Dousset, X. 2000. Detection, purification and partial characterization of a novel bacteriocin substance produced by *Lactococcus lactis* subsp. lactis b14 isolated from Boza-Bulgariantraditional cereal beverage. *Biocatalysis*, 41: 47-53.
- Edward, A.S., Boris, V.E., Vladimir, V.P., Evgeni, V.M., Irina, P.M., Valery, N.B., Vladimir, P.L., Olga, E.S., Yuri, N.K., Yuri, G.S., Gregory, R.S., Bruce, S.S. and Norman, J.S. 2008. Diverse antimicrobial killing by *Enterococcus faecium* E 50-52 bacteriocin. *Journal of Agriculture and Food Chemistry*, 56: 1942–1948.
- Esther, I., Audrey, B., Christine, S., Yimin, C., Eric, M., Alain, V.D. and Sai^ad, E. 2008. Production of Enterocins L50A, L50B, and IT, a new enterocin, by *Enterococcus faecium* IT62, a strain isolated from Italian ryegrass in Japan. *Antimicrobial Agents and Chemotherapy*, 56: 1917–1923.
- Ferreira, A.E., Canal, N., Morales, D., Fuentefria, D.B., and Corcao, G. 2007. Characterization of Enterocins produced by *Enterococcus mundtii* isolated from humans faeces. *Brazilian Archives of Biology and Technology*, 50: 249-258.
- Foulquié Moreno, M.R., Sarantinopoulos, P., Tsakalidou, E. and De Vuyst, L. 2006. The role and application of enterococci in food and health. *International Journal of Food Microbiology*, 106: 1-24.
- Fuller, R. 1998. Probiotics in man and animals. Journal of Applied Bacteriology, 66: 365-378.
- Galvez, A., Valdivia, E., Abriouel, H., Camafeita, E., Mendez, E., Martinez-Bueno, M. and Maqueda, M. 1998. Isolation and characterization of enterocin EJ97, a bacteriocin produced by *Enterococcus faecalis* EJ97. Archives of Microbiology, 171: 59-65.
- Garneau, S., Maartin, N. I. and Vedras, J. C. 2002. Twopeptide bacteriocins produced by lactic acid bacteria. *Biochimie*, 84: 577-592.
- Gilliland, S E. 1996. Special addition culture In: Cogan TM, Accolas JP (eds). *DAIRY starter cultures;* New York: VCH Publishers, 25-46.
- Goldin, B. 1997. Alternation in fecal microflora enzymes related to diet, age, lactobacillus supplements, and dimethylhydrazine, Cancer, 40, 2421-2426.
- Guandalini, S, Pensabene, L, Zikri, M.A., Dias, J.A., Casali L.G., Hoekstra, H, Kolacek, S, Masasar, K, Micetic-Turk, D, Papadopoulou, A, De Sousa, J.S., Sandhu, B, Szajewska, H, Weizman, Z. 2000. *Lactobacillus* GG administered in oral rehydration solution to children with acute diarrhea: a multicentre European trial. *Journal of pediatric Gastroenterology Nutrition*, 30: 54-60.

- Holo, H., Nilssen, T. and Nes, I. F. 1991. Lactococcin A, a new bacteriocin from *Lactococcus lactis subsp. cremoris*: isolation and characterization of the protein and its gene. *Journal of Bacteriology*, 173: 3879-3887.
- John, A., Renye, J., George, A., Somkuti, M. P., Diane, L. and Van, H. 2009. Characterizationof antilisterial bacteriocins produced by *Enterococcus faecium* and *Enterococcus durans* isolates from Hispanic-style cheeses. *Journal of Industrial Microbiology and Biotechnology*, 36: 261–268.
- Kang, J.H. and Lee, M.S. 2005. Characterization of a bacteriocin produced by *Enterococcus faecium* GM-1 isolated from an infant. Klaenhammer, T. R. 1993. Genetics of bacteriocins produced by lactic acid bacteria.*FEMS Microbiology Reviews*, 12: 39-86.
- Koehler, W. 2007. The present state of species within the genera Streptococcus and Enterococcus. International Journal of Medical Microbiology, 297: 133–150.
- Lavermicocca, P., Valeria, F., Evidente, A. Lazzaroni, S, Corsetti, A, Gobbetti, M. 2000. Purification and characteristics of Characterization of noval antifungal compounds by sourdough *Lactobacillus plantarum* 12 B. *Applied and Environmental Microbiology*, 66:4084-4090.
- Lee, N. K. and Paik, H.D. 2001. Partial characterization of lacticin NK24, a newely identified bacteriocin of *Lactococcus lactis* NK24 isolated from Jeot-gal. *Food Microbiology*, 18: 17-24.
- Marteau, PR., De Verse M, Cellier CJ, Schrezenmeir J. 2001. Protection from gastrointestinal disease with the use of probiotics. *American Journal of Clinical Nutition*, 73: 4305-4365.
- Metaxopoulos, J. and Drosinos, E.H. 2002. Characterization of two bacteriocins produced by *Leuconostoc mesenteroides* L124 and *Lactobacillus curvatus* L442, isolated from dry fermented sausages. *World Journal of Microbiology* and *Biotechnology*, 18: 847–856.
- Navarro, L., Zaragaza, M., Saenz, J., Ruiz-Larrea, F. and Torres, C. 2000. Bacteriocin production by lactic acid bacteria isolated from Rioja red wines. *Journal of Applied Microbiology*, 88: 44-51.
- Nes, I. F. and Holo, H. 2000. Class II antimicrobial peptides from lactic acid bacteria. *Biopolymers*, 55: 50-61.
- Ohmomo, S., Murata, S., Katayama, N., Nitisinprasart, S., Kobayashi, M., Nakajima, T., Yajima, M. and Nakanishi, K. 2000. Purification and some characteristics of enterocin ON-157, a bacteriocin produced by *Enterococcus faecium* NIAI 157. *Journal of Applied Microbiology*, 88: 81-89.

- Papagianni, M. and Anastasiadou, S. 2009. Pediocins: The bacteriocins of pediococci. Sources, production, properties and applications. *Microbial Cell Factories*, 8: 1-16.
- Parente, E. and Hill, C. 1992. A comparison of factors affecting the production of two bacteriocins from lactic acid bacteria. *Journal of Applied Bacteriology*, 73: 290-298.
- Piard, J. C. and Desmazeaud, M. 1992. Inhibiting factors produced by lactic acid bacteria: Bacteriocins and other antibacterial substances *Le Lait*, 72 :113-142.
- Salminen S, Deighton MA, Benno Y, Gaback SL 1998a. Lactic acid bacteria in health and disease. In Salminen S, Vonwright A (eds). Lactic acid bacteria:Microbial an functional aspects 2nd Edition. New York: Marcel Dekker Inc, 211-254.
- Shin, M.S., Han, S.K., Ryu, J.S., Kim, K.S. and Lee, W.K. 2008. Isolation and partial characterization of a bacteriocin produced by *Pediococcus pentosaceus* K23-2 isolated from Kimchi. *Journal of Applied Microbiology*, 105: 331-339.
- Simonov'a, M. and Laukov'a, A. 2007. Bacteriocin activity of Enterococci from rabbits. *Veterinary Research Communications*, 31: 143–152.
- Simova, E. D., Beshkova, D.B. and Dimitrov, Z. 2009. Characterization and antimicrobial spectrum of bacteriocins produced by lactic acid bacteria isolated from traditional Bulgarian dairy products. *Journal of Applied Microbiology*, 106: 692-701.
- Tagg, J. R., Dajani, A. S. and Wannamaker, L. W. 1976. Bacteriocins of gram-positive bacteria. *Bacteriological Reviews*, 40: 722-756.
- Tiwari, S. K. and Srivastava, S. 2008. Statistical optimization of culture components for enhanced bacteriocin production by *Lactobacillus plantarum* LR/14. *Food Biotechnology*, 22: 64–77.
- Torodov, S. D. and Dicks, L. M. T. 2005. *Lactobacillus plantarum* isolated from molasses produces bacteriocins active against Gram-negative bacteria. *Enzyme* and *Microbial Technology*, 36: 65-80
- Van Reenen, C.A., L.M.T. Dicks and M.L. Chikindas, 1998. Isolation, purification and partial characterization of plantaricin 423, a bacteriocins produced by *Lactobacillus plantarum. Journal of Applied Microbiology* 84: 1131-1137.
